L Number	Hits	Search Text	DB	Time stamp
1	132	transgene and (e1a with e1b)	USPAT;	2001/11/16 17:43
			US-PGPUB;	
			EPO; JPO;	
:			DERWENT;	
			IBM TDB	
2	127	transgene and (e1a with e1b) and promoter	USPAT;	2001/11/16 17:43
_			US-PGPUB;	
1			EPO; JPO;	
			DERWENT;	
			IBM TDB	
3	316	(e1a with e1b) and promoter	USPAT;	2001/11/16 17:43
	310	(e la with e lb) and promoter	US-PGPUB;	
		•	EPO; JPO;	
			DERWENT;	
			IBM TDB	
4	127	(e1a with e1b) and promoter and transgene	USPAT;	2001/11/16 17:43
			US-PGPUB;	2001/11/10 17:40
j			EPO; JPO; DERWENT:	
5				
			IBM TDB	0004/44/46 47:44
	520	e1a and e1b	USPAT;	2001/11/16 17:44
			US-PGPUB;	
6			EPO; JPO;	
			DERWENT;	
			IBM TDB	
	95	(e1a and e1b) with vector	USPAT;	2001/11/16 17:44
			US-PGPUB;	
7			EPO; JPO;	
			DERWENT;	
			IBM TDB	
	37	((e1a with e1b) and promoter and transgene) and ((e1a and	USPAT;	2001/11/16 17:44
		e1b) with vector)	US-PGPUB;	
ľ		,	EPO; JPO;	
			DERWENT;	
			IBM TDB	
_	0	(AAV and adenovir?) with hybrid	USPAT; IBM	2001/01/31 09:34
	J	, are and additionally many many	TDB	
_	113	(host adj cell?) and rep and cap	USPAT; IBM	2001/01/31 09:36
·	113	(nost adjoen;) and rep and odp	TDB	
	0	((host adj cell?) and rep and cap) and adeno?	USPAT; IBM	2001/01/31 09:36
-	U	((nost adjicent) and repland cap) and adenot	TDB	
	7	((heat adj cells) and rep and can) and ((inducs or cometitutivs)	USPAT; IBM	2001/01/31 09:41
-	7	1 **	TDB	2001/01/01 00:41
i	4.5	with promoter?)		2001/01/31 00:54
-	15	((host adj cell?) and rep and cap) and E1a and E1b and E2a	USPAT; IBM	2001/01/31 09:54
	_	(/b and and and and and and E45 and E45 and E65	TDB	2001/01/21 00:54
-	0	((host adj cell?) and rep and cap) and E1a and E1b and E2a	EPO; JPO;	2001/01/31 09:54
		1. 1. 20. 30.	DERWENT	2004/04/24 00:55
-	254	helper with virus	EPO; JPO;	2001/01/31 09:55
1			DERWENT	0004/04/04 00 50
-	1268	helper with virus	USPAT; IBM	2001/01/31 09:56
			TDB	
-	0	(((host adj cell?) and rep and cap) and E1a and E1b and E2a)	USPAT; IBM	2001/01/31 09:56
		not (helper with virus)	TDB	
-	741	helper adj virus	USPAT; IBM	2001/01/31 09:56
			TDB	
_	0	(((host adj cell?) and rep and cap) and E1a and E1b and E2a)	USPAT; IBM	2001/01/31 09:57
	_	not (helper adj virus)	TDB	
_	3		USPAT; IBM	2001/01/31 09:58
	J	and transgene?	TDB	
_	0	(((host adj cell?) and rep and cap) and E1a and E1b and E2a)	EPO; JPO;	2001/01/31 09:58
	o o	and transgene?	DERWENT	
		and transgene:		
	15	(((host adj cell?) and rep and cap) and E1a and E1b and E2a)	USPAT; IBM	2001/01/31 09:59

-	3	9534670.pn.	USPAT;	2001/11/16 14:01
		·	US-PGPUB;	
			EPO; JPO;	
1	!		DERWENT;	
			IBM TDB	
-	3	9615777.pn.	USPAT;	2001/11/16 14:01
			US-PGPUB;	
			EPO; JPO;	
			DERWENT;	
			IBM TDB	
-	1	6258595.pn.	USPAT; IBM	2001/11/16 17:14
			TDB	
-	70	E1a adj promoter	USPAT;	2001/11/16 17:14
	ļ		US-PGPUB;	
1			EPO; JPO;	
•	1		DERWENT;	
:	1		IBM TDB	
-	20	E2a adj promoter	USPAT;	2001/11/16 17:14
	İ		US-PGPUB;	
			EPO; JPO;	
			DERWENT;	
]		IBM TDB	
-	27	E1b adj promoter	USPAT;	2001/11/16 17:15
	1		US-PGPUB;	
			EPO; JPO;	
	1		DERWENT;	
			IBM TDB	
-	1	(a.a.a.) bronners, and (a.a.a.) bronners, and (a.a.a.)	USPAT;	2001/11/16 17:22
	1	promoter)	US-PGPUB;	
			EPO; JPO;	
			DERWENT;	
	1		IBM TDB	1

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FILE 'MEDLINE' ENTERED AT 16:59:07 ON 16 NOV 2001
L1
            49 S E1A(W)PROMOTER
L2
             43 S E2A(W)PROMOTER
             1 S E2B(W)PROMOTER
L3
             34 S E1B WITH PROMOTER
L4
L5
             0 S L4 AND L1
L6
             29 S E1A AND E1B AND E2A
L7
L8
L9
        84905 S PROMOTER?
         16156 S L7 AND 6
             9 S L7 AND L6
L10
             27 S E1B(W) PROMOTER
             0 S L1 AND L10 AND L2
L11
             0 S L1 AND L10
L12
             0 S L1 AND L2
L13
             0 S L10 AND L2
L14
            1 S HYBRID AAV AND ADENOVIRUS
0 S TRANSGENE AND REP AND CAP AND E1A AND E2A
L15
L16
          131 S TRANSGENE AND AAV
L17
           61 S REP AND CAP
L18
             7 S L17 AND L18
L19
            19 S TRANSGENE AND ITR
L20
L21
             1 S L19 AND L20
L22 29 S E1A AND E1
L23 1273466 S INDUC?
L24 84905 S PROMOTER?
             29 S E1A AND E1B AND E2A
L25
              1 S L22 AND L23 AND L24
L26
             17 S E1A AND CMV
             3 S E2A AND CMV
L27
              0 S L26 AND L27
L28
              3 S HYBRID ADENOVIRUS AND AAV
L29
L30
            30 S E2A AND TRANSGENE
            453 S E1A AND E1B
L31
L32
             0 S L30 AND L31
L33
              0 S L30 AND ITR
                E GAO/AU
                E GAO G/AU
             75 S E3
L34
              E GAO GUANGPING/AU
               E GAO GUANGPING/AU
L35
             2 S HYBRID AND L34
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L18 ANSWER 7 OF 11 MEDLINE

ACCESSION NUMBER: 95134094 MEDLINE

DOCUMENT NUMBER: 95134094 PubMed ID: 7832644

TITLE: Unusual splice sites in the E1A-E1B

cotranscripts synthesized in adenovirus type 40-infected

A549 cells.

AUTHOR: Ishida S; Fujinaga Y; Fujinaga K; Sakamoto N; Hashimoto S

CORPORATE SOURCE: Cancer Research Institute, Sapporo Medical College,

Japan.

SOURCE: ARCHIVES OF VIROLOGY, (1994) 139 (3-4) 389-402.

Journal code: 8L7; 7506870. ISSN: 0304-8608.

PUB. COUNTRY: Austria

Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199502

ENTRY DATE: Entered STN: 19950307

Last Updated on STN: 19970203 Entered Medline: 19950222

The adenovirus El DNA region consists of two transcription units, ElA and ElB. In this paper we report that the ElA-ElB cotranscripts containing sequences of both the ElA and ElB regions are synthesized during adenovirus type 40 (Ad40) infection of A549 cells. Cytoplasmic RNA was isolated from Ad40-infected A549 cells at 24, 72, and 100 h post infection (p.i.). The complementary (c) DNA was synthesized by reverse transcription using an oligo-dT primer and then amplified by the

polymerase chain reaction (PCR) using primers derived from the E1A and

E1B

regions. The cDNAs thus amplified were sequenced either directly or after cloning into bacteriophage M13 ${\bf vectors}$. Analysis of cDNA

indicated that the ${\tt E1A-E1B}$ cotranscripts are

synthesized at 72 h p.i., but not at 24 or 100 h p.i. Nucleotide sequences

of three cDNAs of the E1A-E1B cotranscripts indicated

that the cotranscripts originate from the E1A promoter and lack sequences for both the E1A poly(A) site and E1B cap site. The splices create open reading frames for **E1A-E1B** fused polypeptides around

the E1A-E1B junctions in these mRNAs. Most

interestingly, the sequence analysis showed that the 5' and 3' splice junctions in the two ${\tt E1A-E1B}$ cotranscripts do not

conform to the splice consensus GT-AG rule. Our results thus suggest that factor(s) which lead to unusual splicing in the E1 mRNAs are present in Ad40-infected A549 cells.

L18 ANSWER 3 OF 11 MEDLINE

ACCESSION NUMBER: 1998105729 MEDLINE

DOCUMENT NUMBER: 98105729 PubMed ID: 9444984

TITLE: Production and characterization of improved adenovirus

vectors with the E1, E2b, and E3 genes deleted.
Amalfitano A; Hauser M A; Hu H; Serra D; Begy C R;

Chamberlain J S

CORPORATE SOURCE: Department of Pediatrics, Duke University Medical Center,

Durham, North Carolina 27710, USA.. amalf001@mc.duke.edu

SOURCE: JOURNAL OF VIROLOGY, (1998 Feb) 72 (2) 926-33.

Journal code: KCV; 0113724. ISSN: 0022-538X.

PUB. COUNTRY: United States

Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

AUTHOR:

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199802

ENTRY DATE: Entered STN: 19980226

Last Updated on STN: 19980226 Entered Medline: 19980218

Adenovirus (Ad)-based vectors have great potential for use in AB the gene therapy of multiple diseases, both genetic and nongenetic. While capable of transducing both dividing and quiescent cells efficiently, Ad vectors have been limited by a number of problems. Most Ad vectors are engineered such that a transgene replaces the Ad Ela, Elb, and E3 genes; subsequently the replication-defective vector can be propagated only in human 293 cells that supply the deleted El gene functions in trans. Unfortunately, the use of high titers of El-deleted vectors has been repeatedly demonstrated to result in low-level expression of viral genes still resident in the vector. In addition, the generation of replication-competent Ad (RCA) by recombination events with the E1 sequences residing in 293 cells further limits the usefulness of E1-deleted Ad vectors. We addressed these problems by isolating new Ad vectors deleted for the E1, E3, and the E2b gene functions. The new vectors can be readily grown to high titers and have several improvements, including an increased carrying capacity and a theoretically decreased risk for generating RCA. We have also demonstrated that the further block to Ad vector replication afforded by the deletion of both the El and E2b genes significantly diminished Ad late gene expression in comparison to a conventional E1-deleted vector, without destabilization of the modified vector genome. The results suggested that these modified vectors may be very useful both for in vitro and in vivo gene therapy applications.

ACCESSION NUMBER:

83307850 MEDLINE

DOCUMENT NUMBER:

83307850 PubMed ID: 6615164

TITLE:

Adenoviruses and human tumors: regulation of eukaryotic

chromatin structure?.

AUTHOR:

Berencsi G; Nasz I

SOURCE:

ARCHIV FUR GESCHWULSTFORSCHUNG, (1983) 53 (3) 239-52.

Journal code: 746; 0372411. ISSN: 0003-911X.

PUB. COUNTRY:

GERMANY, EAST: German Democratic Republic

Journal; Article; (JOURNAL ARTICLE)

LANGUAGE:

English

FILE SEGMENT:

Priority Journals

ENTRY MONTH:

198310

ENTRY DATE:

Entered STN: 19900319

Last Updated on STN: 19900319

Entered Medline: 19831021

AB Adenoviruses possess four early gene clusters. **E1A**, and **E1B** sequences are directly involved in the initiation of

transformation of both rodent, and human cells. Oncogenic potential of

the

in vitro immortalized rodent cells is also dependent on the expression of E1A, and E1B. Genes E2A, and E2B are

coding for DNA-binding proteins. These regions may influence the

frequency

of in vitro transformation if complete virions were used for the immortalization of rodent cells. The existence of the E3 gene cluster, which has been shown to be non-essential to the productive replication in certain host cells, is unique for the adenovirus family. Foreign DNA fragments inserted into E3 or replacing it may modify the host range of adenovirus, and may initiate oncogenic transformation of rodent cells.

May

cellular oncogenes replace the non-essential region of the adenoviral genome? Are host-range modifications, the formation of defective genomes, and interactions of viral, and cellular DNA in vivo biological properties of adenoviruses? These questions are discussed in the light of recent findings concerning virus-coded functions which may modify chromatin structure, and may be associated with oncogenic potential of adenoviruses in the natural hosts.

L21 ANSWER 12 OF 33 MEDLINE

ACCESSION NUMBER: 1998285701 MEDLINE

DOCUMENT NUMBER: 98285701 PubMed ID: 9621003
TITLE: Characterization of wild-type

adeno-associated virus type 2-like particles generated

during recombinant viral vector production and

strategies for their elimination.

AUTHOR: Wang X S; Khuntirat B; Qing K; Ponnazhagan S; Kube D M;

Zhou S; Dwarki V J; Srivastava A

CORPORATE SOURCE: Department of Microbiology and Immunology, Indiana

University School of Medicine, Indianapolis, Indiana

46202,

USA.

CONTRACT NUMBER: HL-48342 (NHLBI)

HL-53586 (NHLBI) HL-58881 (NHLBI)

+

SOURCE: JOURNAL OF VIROLOGY, (1998 Jul) 72 (7) 5472-80.

Journal code: KCV; 0113724. ISSN: 0022-538X.

PUB. COUNTRY: United States

Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199807

ENTRY DATE: Entered STN: 19980713

Last Updated on STN: 19980713 Entered Medline: 19980701

The pSub201-pAAV/Ad plasmid cotransfection system was developed to AB eliminate homologous recombination which leads to generation of the wild-type (wt) adeno-associated virus type 2 (AAV) during recombinant vector production. The extent of contamination with wt AAV has been documented to range between 0.01 and 10%. However, the precise mechanism of generation of the contaminating wt AAV remains unclear. To characterize the wt AAV genomes, recombinant viral stocks were used to infect human 293 cells in the presence of adenovirus. Southern blot analyses of viral replicative DNA intermediates revealed that the contaminating AAV genomes were not authentic wt but rather wt AAV-like sequences derived from recombination between (i) AAV inverted terminal repeats (ITRs) in the recombinant plasmid and (ii) AAV sequences in the helper plasmid. Replicative AAV DNA fragments, isolated following amplification through four successive rounds of amplification in adenovirus-infected 293 cells, were molecularly cloned and subjected to nucleotide sequencing to identify the **recombinant** junctions. Following sequence analyses of 31 different ends of AAV-like genomes derived from two different recombinant vector stocks, we observed that all recombination events involved 10 nucleotides in the AAV D sequence distal to viral hairpin structures. We have recently

documented that the first 10 nucleotides in the D sequence proximal to the

AAV hairpin structures are essential for successful replication and encapsidation of the viral genome (X.-S. Wang et al., J. Virol. 71:3077-3082, 1997), and it was noteworthy that in each recombinant junction sequenced, the same 10 nucleotides were retained. We also observed that adenovirus ITRs in the helper plasmid were involved in illegitimate recombination with AAV ITRs, deletions of which significantly reduced the extent of wt AAV-like particles. Furthermore, the combined use of recombinant AAV plasmids lacking the distal 10 nucleotides in the D sequence and helper

L18 ANSWER 11 OF 11 MEDLINE

ACCESSION NUMBER: 88275039 MEDLINE

DOCUMENT NUMBER: 88275039 PubMed ID: 3292790

TITLE: High-level eucaryotic in vivo expression of biologically

active measles virus hemagglutinin by using an adenovirus

type 5 helper-free vector system.

AUTHOR: Alkhatib G; Briedis D J

CORPORATE SOURCE: Department of Microbiology and Immunology, McGill

University, Montreal, Quebec, Canada.

SOURCE: JOURNAL OF VIROLOGY, (1988 Aug) 62 (8) 2718-27.

Journal code: KCV; 0113724. ISSN: 0022-538X.

PUB. COUNTRY: United States

Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 198808

ENTRY DATE: Entered STN: 19900308

Last Updated on STN: 19900308 Entered Medline: 19880819

AB The entire measles virus (MV) hemagglutinin (HA)-coding region was

reconstructed from cloned cDNAs and used as part of a hybrid

transcription

unit to replace a region of the adenovirus type 5 genome corresponding to the entire Ela transcription unit and most of the Elb transcription unit. The resulting recombinant virus was stable and able to replicate to high titers in 293 cells (which constitutively express the complementary

Ela-Elb functions) in the absence of helper virus. During infection of 293 cells, the hybrid virus expressed MV HA protein which was indistinguishable from that expressed in MV-infected cells in terms of immunoreactivity, gel mobility, glycosylation, subcellular localization, and biologic activity. Infection of 293 cells with the hybrid virus led to high-level synthesis of the MV HA protein (equivalent

to 65 to 130% of the level seen in MV-infected cells). At late times

after

high-multiplicity hybrid virus infection of HeLa and Vero cells (which do not express E1 functions), the level of HA protein synthesis was at least 35% of that seen in 293 cells. This MV-adenovirus recombinant will be useful in the study of the biologic properties of the MV HA protein and

in

assessment of the potential usefulness of hybrid adenoviruses as live-virus vaccine **vectors**.